Novel Pyridyl Ring C5 Substituted Analogues of Epibatidine and 3-(1-Methyl-2(S)pyrrolidinylmethoxy)pyridine (A-84543) as Highly Selective Agents for Neuronal Nicotinic Acetylcholine Receptors Containing $\beta 2$ Subunits

Zhi-Liang Wei,[†] Yingxian Xiao,[‡] Hongbin Yuan,[†] Maryna Baydyuk,[‡] Pavel A. Petukhov,[†] John L. Musachio,[§] Kenneth J. Kellar,[‡] and Alan P. Kozikowski^{*,†}

Department of Medicinal Chemistry and Pharmacognosy, College of Pharmacy, University of Illinois at Chicago, 833 South Wood Street, Chicago, Illinois 60612, Department of Pharmacology, Georgetown University, 3900 Reservoir Road, NW, Washington, D.C. 20057, and Department of Radiology, Johns Hopkins University School of Medicine, Baltimore, Maryland 22287

Received September 16, 2004

Abstract: Introduction of a hydrophobic or hydrogen-bonding alkynyl group into the C5 position of the pyridyl ring of epibatidine and A-84543 significantly increased the selectivity for neuronal nicotinic acetylcholine receptors (nAChRs) containing $\beta 2$ subunits over nAChRs containing $\beta 4$ subunits (K_i ratio up to 92000-fold). Our data indicate that the extracellular domains of the nAChRs are sufficiently different to allow for the design of novel ligands with high affinity and selectivity for the nAChR subtypes.

Neuronal nicotinic acetylcholine receptors (nAChRs) are ligand-gated ion channels.¹ These receptors hold considerable promise as therapeutic targets for the treatment of disorders of the central nervous system (CNS) and peripheral nervous system.² Most nAChRs are heteromeric pentamers. The subunits that comprise the receptors have a common general structure consisting of a large extracellular N-terminal domain that contains the binding sites for acetylcholine, nicotine, and other ligands, four membrane-spanning hydrophobic segments, and a large intracellular domain between the third and the fourth membrane-spanning segments.³ Twelve neuronal nAChR subunits ($\alpha 2-\alpha 10$ and $\beta 2-\beta 4$) have been identified in vertebrates, and different combinations of these subunits define nAChR subtypes.⁴

Subtype selectivity is a critical issue for the effectiveness and safety of drugs. To selectively affect different physiological functions pharmacologically, it is very important to have drugs that preferentially act on specific receptor subtypes. Similarly, nAChR subtypeselective ligands may prove useful in the diagnosis of brain pathology by means of positron emission tomography (PET) imaging.⁵ However, finding compounds that discriminate among nAChR subtypes has proven to be difficult because of the large number of potential

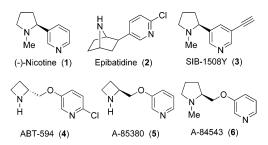


Figure 1.

receptor subtypes and the relatively subtle differences in their structures. There are a large number of nicotinic agonists and noncompetitive antagonists (channel blockers), but very few of these nicotinic compounds are subtype-selective.⁶

Development of selective agonists or antagonists may therefore result in new and potentially useful therapeutic agents. One of the important targets for selective drugs is the $\alpha 4\beta 2$ nAChR subtype, which is the most abundant nAChRs in the brain. Nicotine (1) and epibatiline (2), as naturally occurring agonists of nAChRs, have attracted interest as starting points for modification leading to structures with improved pharmacological properties.⁶⁻⁸ In particular, within the past several years, a series of pyridyl ether compounds, including ABT-594 (4), A-85380 (5), and A-84543 (6), were synthesized as high-affinity chain-extended analogues of nicotine (1) (Figure 1).⁹ Although some of these compounds are selective, their discrimination among receptor subtypes is generally about 1 or 2 orders of magnitude.¹⁰ Among the new nicotinic compounds developed in recent years, A-85380 and one of its analogue, 5-iodo-A-85380, showed the highest selectivities between the $\alpha 4\beta 2$ subtype and $\alpha 3\beta 4$ subtype, the main ganglionic nAChR population.¹¹ We found previously that some modifications at the C5 position of the pyridyl ring of 5 significantly enhanced the selectivity to 4 orders of magnitude between $\alpha 4\beta 2$ and $\alpha 3\beta 4$ subtypes.¹² In continuation of our efforts to find selective neuronal nAChR ligands,¹³ we report here our discovery that introduction of hydrophobic or hydrogen-bonding alkynyl substituents at the C5 position of the pyridyl ring of 6 and dechloroepibatidine imparts very high selectivity for receptors containing $\beta 2$ subunits over receptors containing $\beta 4$ subunits.

The C5 position of the pyridyl moiety of A-84543 tolerated sterically bulky substituents without losing its binding affinity at $\alpha 4\beta 2$ nAChR.⁹ Our aim therefore was to investigate the effects of C5 substituents of the pyridine ring on the binding affinity and subtype selectivity at neuronal nAChRs caused by steric factors and the hydrophobicity of the newly introduced group. Introduction of an ethynyl substituent at the C5 position of the pyridyl ring of 1 led to SIB-1508Y (3) with altered subtype selectivity for neuronal nAChRs.¹⁴ Thus, 5-alkynyl-substituted A-84543 analogues 8-15 were prepared (Scheme 1) and evaluated by binding assays at six heterologously expressed nAChR subtypes ($\alpha 2\beta 2$, $\alpha 2\beta 4$, $\alpha 3\beta 2$, $\alpha 3\beta 4$, $\alpha 4\beta 2$, and $\alpha 4\beta 4$). In particular, we compared the affinities of these ligands at the $\alpha 3\beta 4$ subtype with their affinities at the $\alpha 4\beta 2$ subtype. The

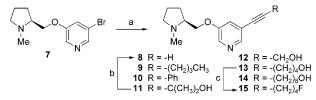
^{*} To whom correspondence should be addressed. Phone: 312-996-7577. Fax: 312-996-7107. E-mail: kozikowa@uic.edu.

[†] University of Illinois at Chicago.

[‡] Georgetown University.

[§] Johns Hopkins University. Present address: NIMH, National Institutes of Health, 9000 Rockville Pike, Bethesda, Maryland 20892.

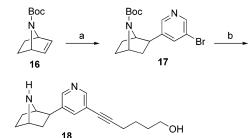
Scheme 1^a



^a Reagents: (a) alkyne, 10% Pd–C (cat.), CuI (cat.), K₂CO₃, DME, H₂O, reflux, 72 h, 55–95%; (b) NaH (cat.), toluene, 120 °C, 1 h, 99%; (c) (i) I₂, PPh₃, imidazole, CH₂Cl₂, 92%; (ii) AgF, acetonitrile, 10 h, 57%.

results are summarized in Table 1. The $\alpha 3\beta 4$ subtype is found in many sympathetic ganglia, while the $\alpha 4\beta 2$ subtype is the predominant nAChR in the forebrain; therefore, the affinity ratios of drugs at these subtypes can help to predict the likelihood of autonomic nervous system side effects of drugs aimed at the predominant receptor subtype in the forebrain.¹¹ Neither nicotine (1)nor epibatidine (2) shows significant selectivity among the six nAChR subtypes (<45-fold). A-84543 (6) possesses very high affinity for all three nAChR subtypes containing $\beta 2$ subunits but much lower affinity for the subtypes containing $\beta 4$ counterparts, although the highest selectivity among the six nAChR subtypes is still less than 750-fold. The improved selectivity suggests the possibility of developing subtype-selective ligands. Introduction of additional substituent groups at the C5 position of the pyridyl ring of 6 resulting in 8-15 did not cause marked differences in the binding affinities at the $\alpha 4\beta 2$ -containing subtype (within ~4-fold). However, it is noteworthy that introduction of a 5-alkynyl group decreased the affinities of 8–15 at the $\alpha 3\beta 4$ subtype, in some cases markedly, resulting in improved selectivity between the nAChR subtypes. As expected, introduction of an ethynyl substituent at the C5 position of the pyridyl ring of 6 led to 8 with improved nAChR subtype selectivity between $\alpha 4\beta 2$ and $\alpha 3\beta 4$ (>4000-fold). Extension of the ethynyl substituent with hydrophobic groups or hydrogen-bonding groups improved the selectivity (as shown by 9-14). The 6-hydroxy-1-hexynyl substituent at the C5 position of the pyridine ring of **13** is a preferred group for attaining the expected high binding affinity at the $\alpha 4\beta 2$ receptor (0.85 nM) and very high subtype-selectivity (>74000-fold). Similarly the fluoride-containing analogue 15 possesses much higher





 a Reagents: (a) 3,5-dibromopyridine, Pd(PPh_3)_4 (cat.), piperidine, HCO_2H, DMF, 80 °C, 72 h, 61%; (b) (i) 6-[(tert-butyldimethyl-silyl)oxy]-1-hexyne, Pd(PPh_3)_2Cl_2 (cat.), CuI (cat.), Bu_4NI, Et_3N, DMF, reflux, 48 h, 93%; (ii) CF_3CO_2H, CH_2Cl_2, 90%.

affinities at receptors composed of an α subunit in combination with the $\beta 2$ subunit than with the $\beta 4$ subunit.

If epibatidine **2** and the 3-pyridyl ether **6** bind at each nAChR in a common manner, then the significant improvement of the subtype selectivity of 13 that is achieved by introducing a 6-hydroxy-1-hexynyl group at the C5 position of the pyridyl ring should also apply to the corresponding epibatidine analogues. A very recent study reveals that introduction of a bulky phenyl group at the C5 position of the pyridyl ring of epibatidine results in ligands that are antagonists at nAChRs.¹⁵ On the other hand, it has been shown that dechloroepibatidine binds with similar affinity as epibatidine at the $\alpha 4\beta 2$ nAChR subtype.¹⁶ Thus, 5-(6hydroxy-1-hexynyl)-substituted dechloroepibatidine analogue 18 was prepared by using the reductive Heck reaction of 16^{16a} with 3,5-dibromopyridine followed by the Sonogashira reaction as the key steps (Scheme 2). As shown in Table 1, 18 also possesses subnanomolar affinities at the β 2-containing subtypes, although its affinities are 2- to 8-fold lower than those of epibatidine at each of the β 2-containing subtype. It is noteworthy that **18** was quite selective for an α subunit paired with the $\beta 2$ versus the $\beta 4$ subunit. Thus, the $\alpha 3\beta 4/\alpha 4\beta 2$ affinity ratio of 18 was >2385, while epibatidine, which itself binds to most nAChR subtypes with picomolar affinity, displays a ratio of only \sim 9.

To further explore the binding modes of these ligands at different nicotinic receptor subtypes and to uncover the underlying mechanism for the observed selectivity,

Table 1. Binding Affinities of (-)-Nicotine (1), (±)-Epibatidine (2), A-84543 (6), 8-15, and 18 at Six nAChR Subtypes^a

	binding affinity K_{i} (nM)						selectivity	
ligand	$\alpha 2\beta 2$	$\alpha 2\beta 4$	$\alpha 3\beta 2$	$\alpha 3\beta 4$	$\alpha 4\beta 2$	$\alpha 4\beta 4$	$\alpha 3\beta 4/\alpha 4\dot{\beta 2}$	$cLogP^{b}$
1	12 ± 2	112 ± 21	47 ± 11	443 ± 60	10 ± 2	40 ± 6	44	0.88
2	0.025 ± 0.001	0.095 ± 0.017	0.035 ± 0.011	0.57 ± 0.12	0.061 ± 0.009	0.16 ± 0.01	9	1.55
6	2.5 ± 1.3	320 ± 60	7.7 ± 0.7	1400 ± 400	1.9 ± 0.7	220 ± 70	737	1.83
8	2.5 ± 1.7	750 ± 200	8.2 ± 1.3	6800 ± 2300	1.6 ± 0.7	710 ± 190	4250	2.10
9	9.4 ± 3.8	1900 ± 200	21 ± 1	40000 ± 30000	1.4 ± 0.6	1900 ± 600	28571	4.08
10	4.4 ± 1.9	500 ± 70	8.6 ± 2.1	8100 ± 5700	0.51 ± 0.21	530 ± 160	15882	4.47
11	1.9 ± 0.1	8400 ± 1300	23 ± 2	61000 ± 21000	2.9 ± 1.5	7000 ± 1200	21034	1.35
12	3.4 ± 2.1	1900 ± 100	7.1 ± 0.3	23000 ± 6000	0.93 ± 0.22	1300 ± 300	24731	0.64
13	1.9 ± 0.7	3600 ± 300	13 ± 3	63000 ± 12000	0.85 ± 0.19	1200 ± 100	74118	2.23
14	22 ± 8	$6{,}200\pm800$	48 ± 3	$66,000 \pm 7,000$	6.1 ± 0.5	$2,\!400\pm 600$	10,820	4.34
15	4.2 ± 2.2	$3,700\pm700$	14 ± 1	$88,000 \pm 58,000$	0.95 ± 0.57	$2,000 \pm 1,100$	92,632	3.49
18	0.19 ± 0.09	28 ± 9	0.23 ± 0.09	310 ± 60	0.13 ± 0.03	10 ± 3	2,385	1.15

^{*a*} The K_d values (nM) for [³H]-epibatidine used for calculating K_i values were 0.02 for $\alpha 2\beta 2$, 0.08 for $\alpha 2\beta 4$, 0.03 for $\alpha 3\beta 2$, 0.30 for $\alpha 3\beta 4$, 0.04 for $\alpha 4\beta 2$, and 0.09 for $\alpha 4\beta 4$.¹¹ The K_i values of (–)-nicotine (1) and (±)-epibatidine (2) were published previously¹¹ and are shown here for comparison. The K_i values of **6**, **8–15**, and **18** shown are the mean of three independent measurements. ^{*b*} http://www.daylight.com/daycgi/clogp.

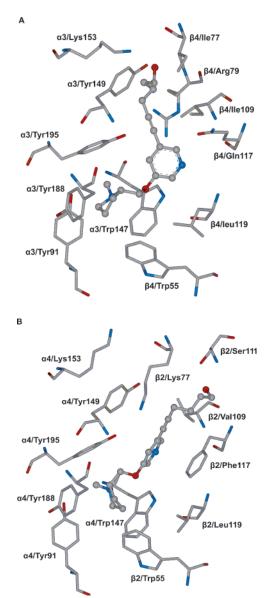


Figure 2. Difference in the binding modes of ligand **13** docked to the models of $\alpha 3\beta 4$ (A) and $\alpha 4\beta 2$ (B) nAChR receptors.

we used the program Autodock3.0¹⁷ to dock these ligands in the rat nicotinic receptor models $\alpha 3\beta 4$ (10LJ) and $\alpha 4\beta 2$ (10LE),¹⁸ which are based on the X-ray structure (119B) of the acetylcholine-binding protein (AChBP).¹⁹ The Phe¹¹⁷ in the $\beta 2$ subunit protein may be involved in the high-affinity binding of nicotine.^{18,20} The docking results confirm that the pyridyl ring and the 5-substitued hydrophobic group of ligand 13 have $\pi - \pi$ and strong hydrophobic interactions with the phenyl ring of Phe¹¹⁷ in the $\alpha 4\beta 2$ protein (Figure 2B), whereas similar lipophilic interactions are not possible with the polar side chain of Gln^{117} in the $\alpha 3\beta 4$ nicotinic receptor (Figure 2A) and other β 4-containing receptors. According to the docking experiment, the pyridyl ring of ligand 13 tends to rotate about 90° to avoid unfavorable interactions with the polar side chain of Gln¹¹⁷. This spatial reorganization does not lead to formation of new hydrogen bonds; thus, the loss of hydrophobic interactions is not balanced, which ultimately leads to weaker binding of ligand 13 in this series to the receptors containing the $\beta 4$ subunits. Overall, this difference in the binding modes may explain the fact that ligands

8–15 in general have much higher affinities for the β 2-containing receptors than for the β 4-containing receptors.

On the basis of their binding affinities and the results from modeling, we conclude that nicotine, epibatidine, and the 3-pyridyl ether analogues bind in a similar fashion at each nAChR subtype. These selective analogues containing appropriately functionalized side chain appendages are quite interesting because, in addition to their general use as pharmacological tools, they may be used in creating fluorescent probes and affinity columns for certain nAChR subtypes. In addition, if labeled with ¹¹C or ¹⁸F, some of these selective ligands could be useful as PET imaging probes.²¹ We have begun studies of the pharmacological actions of some of these ligands at nAChR subtypes to determine whether they are agonists, partial agonists, or antagonists. Moreover, in light of the high affinity and selectivity found for some of these ligands (e.g., 11-13 and 15), efforts to explore them in brain PET studies are underway.

In summary, we have synthesized high-affinity nAChR ligands that display selectivity for nAChRs containing the $\beta 2$ subunit in general and the $\alpha 4\beta 2$ subtype in particular. These studies thus show that the extracellular domains of the nAChRs are sufficiently different to allow for the design of novel ligands with high affinity and selectivity for the nAChR subtypes. Because of the recent advance in solving the structure of the molluscan AChBP¹⁹ and the resulting homology modeling of the extracellular domains of some nAChRs,^{18,20,22} strategies for developing new subtype-selective ligands should become more rational.

Acknowledgment. This work was supported by the National Institutes of Health (Grant R01 DA017980) and by National Institute of Mental Health through the Psychoactive Drug Screening Program (Grant NO1MH32004).

Supporting Information Available: Detailed experimental procedures with spectroscopic data and table of combustion data. This material is available free of charge via the Internet at http://pubs.acs.org.

References

- Karlin, A. Emerging structure of the nicotinic acetylcholine receptors. Nat. Rev. Neurosci. 2002, 3, 102-114.
- (2) Lloyd, G. K.; Williams, M. Neuronal nicotinic acetylcholine receptors as novel drug targets. J. Pharmacol. Exp. Ther. 2000, 292, 461-467.
- (3) Grutter, T.; Changeux, J.-P. Trends Biochem. Sci. 2001, 26, 469.
- (4) Millar, N. S. Assembly and subunit diversity of nicotinic acetylcholine receptors. *Biochem. Soc. Trans.* 2003, 31, 869– 874.
- (5) (a) Fan, H.; Scheffel, U. A.; Rauseo, P.; Xiao, Y.; Dogan, A. S.; Yokoi, F.; Hilton, J.; Kellar, K. J.; Wong, D. F.; Musachio, J. L. [^{125/123}1] 5-Iodo-3-pyridyl ethers: Synthesis and binding to neuronal nicotinic acetylcholine receptors. *Nucl. Med. Biol.* 2001, 28, 911-921. (b) Ding, Y.-S.; Liu, N.; Wang, T.; Marecek, J.; Garza, V.; Ojima, I.; Fowler, J. S. Synthesis and evaluation of 6-[¹⁸F]fluoro-3-(2(S)-azetidinylmethoxy)pyridine as a PET tracer for nicotinic acetylcholine receptors. *Nucl. Med. Biol.* 2000, 27, 381-389. (c) Scheffel, U.; Horti, A. G.; Koren, A. O.; Ravert, H. T.; Banta, J. P.; Finley, P. A.; London, E. D.; Dannals, R. F. 6-[¹⁸F]Fluoro-A-85380: An in vivo tracer for the nicotinic acetylcholine receptor. *Nucl. Med. Biol.* 2000, 27, 51-56.
- (6) Holladay, M. W.; Dart, M. J.; Lynch, J. K. Neuronal nicotinic acetylcholine receptors as targets for drug discovery. J. Med. Chem. 1997, 40, 4169-4194.

- (7) Ferretti, G.; Dukat, M.; Giannella, M.; Piergentili, A.; Pigini, M.; Quaglia, W.; Damaj, M. I.; Martin, B. R.; Glennon, R. A. Binding of nicotine and homoazanicotine analogues at neuronal nicotinic acetylcholinergic (nACh) receptors. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 733–735.
- (8) (a) Tønder, J. E.; Olesen, P. H. Agonists at the α4β2 nicotinic acetylcholine receptors: structure-activity relationships and molecular modelling. *Curr. Med. Chem.* **2001**, *8*, 651-674. (b) Carroll, F. I.; Lee, J. R.; Navarro, H. A.; Ma, W.; Brieaddy, L. E.; Abraham, P.; Damaj, M. I.; Martin, B. R. Synthesis, nicotinic acetylcholine receptor binding, and antinociceptive properties of 2-exo-2-(2',3'-disubstituted 5'-pyridinyl)-7-azabicyclo[2.2.1]heptanes: epibatidine analogues. *J. Med. Chem.* **2002**, *45*, 4755-4761.
- (9) (a) Lin, N.-H.; Li, Y.; He, Y.; Holladay, M. W.; Kuntzweiler, T.; Anderson, D. J.; Campbell, J. E.; Arneric, S. P. Synthesis and structure-activity relationships of 5-substituted pyridine analogues of 3-[2-((S)-pyrrolidinyl)methoxy]pyridine, A-84543: A potent nicotinic receptor ligand. *Bioorg. Med. Chem. Lett.* 2001, 11, 631-633. (b) Lin, N.-H.; Gunn, D. E.; Li, Y.; He, Y.; Bai, H.; Ryther, K. B.; Kuntzweiler, T.; Donnelly-Roberts, D. L.; Anderson, D. J.; Campbell, J. E.; Sullivan, J. P.; Arneric, S. P.; Holladay, M. W. Synthesis and structure-activity relationships of pyridine-modified analogs of 3-[2-((S)-pyrrolidinyl)methoxy]pyridine, A-84543, a potent nicotinic acetylcholine receptor agonist. *Bioorg. Med. Chem. Lett.* 1998, 8, 249-252.
- (10) Gotti, C.; Carbonnelle, E.; Moretti, M.; Zwart, R.; Clementi, F. Drugs selective for nicotinic receptor subtypes: a real possibility or a dream? *Behav. Brain Res.* 2000, *113*, 183–192.
- (11) Xiao, Y.; Kellar, K. J. The comparative pharmacology and up-regulation of rat neuronal nicotinic receptor subtype binding sites stably expressed in transfected mammalian cells. *J. Pharmacol. Exp. Ther.* 2004, *310*, 98–107.
 (12) Kellar, J. K.; Xiao, Y.; Hernandez, S. C.; Perry, D. C. Compara-
- (12) Kellar, J. K.; Xiao, Y.; Hernandez, S. C.; Perry, D. C. Comparative pharmacology and distribution of heteromeric neuronal nicotinic receptors. Abstracts in Program Book, Neuronal Nicotinic Receptors and Ligands: Targets for Medication; NIDA, NIH: Bal Harbour, FL, 2003.
- (13) (a) Wei, Z.-L.; Petukhov, P. A.; Xiao, Y.; Tückmantel, W.; George, C.; Kellar, K. J.; Kozikowski, A. P. Synthesis, nicotinic acetyl-choline receptor binding affinities, and molecular modeling of constrained epibatidine analogues. J. Med. Chem. 2003, 46, 921-924. (b) Wei, Z.-L.; Xiao, Y.; George, C.; Kellar, K. J.; Kozikowski, A. P. Functionalization of the alicyclic skeleton of epibatidine: synthesis and nicotinic acetylcholine receptor binding affinities of epibatidine analogues. Org. Biomol. Chem. 2003, 1, 3878-3881. (c) Wei, Z.-L.; Xiao, Y.; Kellar, K. J.; Kozikowski, A. P. Synthesis and pharmacological characterization of bivalent ligands of epibatidine at neuronal nicotinic acetylcholine receptors. Bioorg. Med. Chem. Lett. 2004, 14, 1855-1858.
- (14) Cosford, N. D. P.; Bleicher, L.; Herbaut, A.; McCallum, J. S.; Vernier, J.-M.; Dawson, H.; Whitten, J. P.; Adams, P.; Chavez-Noriega, L.; Correa, L. D.; Crona, J. H.; Mahaffy, L. S.;

Menzaghi, F.; Rao, T. S.; Reid, R.; Sacaan, A. I.; Santori, E.; Stauderman, K. A.; Whelan, K.; Lloyd, G. K.; McDonald, I. A. (S)-(-)-5-Ethynyl-3-(1-methyl-2-pyrrolidinyl)pyridine Maleate (SIB-1508Y): A novel anti-Parkinsonian agent with selectivity for neuronal nicotinic acetylcholine receptors. J. Med. Chem. **1996**, 39, 3235–3237.

- (15) Carroll, F. I.; Lee, J. R.; Navarro, H. A.; Brieaddy, L. E.; Abraham, P.; Damaj, M. I.; Martin, B. R. Synthesis, nicotinic acetylcholine receptor binding, and antinociceptive properties of 2-exo-2-(2'-substituted-3'-phenyl-5'-pyridinyl)-7-azabicyclo-[2.2.1]heptanes. Novel nicotinic antagonists. J. Med. Chem. 2001, 44, 4039-4041.
- (16) (a) Carroll, F. I.; Liang, F.; Navarro, H. A.; Brieaddy, L. E.; Abraham, P.; Damaj, M. I.; Martin, B. R. Synthesis, nicotinic acetylcholine receptor binding, and antinociceptive properties of 2-exo-2-(2'-substituted 5'-pyridinyl)-7-azabicyclo[2.2.1]heptanes. Epibatidine analogues. J. Med. Chem. 2001, 44, 2229-2237. (b) Badio, B.; Daly, J. W. Epibatidine, a potent analgetic and nicotinic agonist. Mol. Pharmacol. 1994, 45, 563-569.
- (17) Morris, G. M.; Goodsell, D. S.; Halliday, R. S.; Huey, R.; Hart, W. E.; Belew, R. K.; Olson, A. J. Automated docking using a Lamarckian genetic algorithm and empirical binding free energy function. J. Comput. Chem. **1998**, 19, 1639–1662.
- (18) Le Novère, N.; Grutter, T.; Changeux, J.-P. Models of the extracellular domain of the nicotinic receptors and of agonistand Ca²⁺-binding sites. *Proc. Natl. Acad. Sci. U.S.A.* 2002, 99, 3210–3215.
- (19) Brejc, K.; van Dijk, W. J.; Klaassen, R. V.; Schuurmans, M.; van der Oost, J.; Smit, A. B.; Sixma, T. K. Crystal structure of an ACh-binding protein reveals the ligand-binding domain of nicotinic receptors. *Nature* **2001**, *411*, 269–276.
- (20) Costa, V.; Nistri, A.; Cavalli, A.; Carloni, P. A structural model of agonist binding to the α3β4 neuronal nicotinic receptor. Br. J. Pharmacol. 2003, 140, 921–931.
- (21) Zhang, Y.; Pavlova, O. A.; Chefer, S. I.; Hall, A. W.; Kurian, V.; Brown, L. L.; Kimes, A. S.; Mukhin, A. G.; Horti, A. G. 5-Substituted derivatives of 6-halogeno-3-((2-(S)-azetidinyl)methoxy)pyridine and 6-halogeno-3-((2-(S)-pyrrolidinyl)methoxy)pyridine with low picomolar affinity for α4β2 nicotinic acetylcholine receptor and wide range of lipophilicity: Potential probes for imaging with positron emission tomography. J. Med. Chem. 2004, 47, 2453-2465.
- (22) (a) Schapira, M.; Abagyan, R.; Totrov, M. Structural model of nicotinic acetylcholine receptor isotypes bound to acetylcholine and nicotine. *BMC Struct. Biol.* **2002**, *2*, 1. (b) Dutertre, S.; Lewis, R. J. Computational approaches to understand α-conotoxin interactions at neuronal nicotinic receptors. *Eur. J. Biochem.* **2004**, *271*, 2327–2334.

JM0492406